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### Structural filter is defined by a set of interactions:

- typically observed in available structures of protein-ligand complexes;
- > considered to play a crucial role in ligand binding





https://biokinet.belozersky.msu.ru/vsfilt.

#### pubs.acs.org/jcim

#### Application Note

#### vsFilt: A Tool to Improve Virtual Screening by Structural Filtration of Docking Poses

Irina V. Gushchina,<sup>§</sup> Aleksandra M. Polenova, Dmitry A. Suplatov, Vytas K. Švedas, and Dmitry K. Nilov<sup>\*,8</sup>



Gushchina et al. J. Chem. Inf. Mod. (2020) 60, 3692-2696







- hydrogen bonds;
- halogen bonds;
- ➤ ionic interactions;
- >hydrophobic contacts;
- $>\pi$ -stacking;
- $\succ$  cation- $\pi$  interactions



- hydrogen bonds;
- halogen bonds;
- ➤ ionic interactions;
- hydrophobic contacts;
- $>\pi$ -stacking;
- $\succ$  cation- $\pi$  interactions

### Analogs:

- ➤ in-house scripts;
- AutoDock/Raccoon (free);
- Schrödinger/Glide (commercial);
- nAPOLI server





Figure 1. Example of control data used by vsFilt for structural filtration.



automatically identifies atoms involved in the interaction;

> sets the corresponding criteria for filtering



Figure 1. Example of control data used by vsFilt for structural filtration.





recognizes the type of interaction (H-bonding);

- identifies interacting atoms (H-bond donors and acceptors) and their coordinates;
- applies the corresponding distance and angle criteria for structural filtration

#### Table 1. Ligand Functional Groups That Can Be Involved in Structural Filtration with vsFilt, and Their Role in the Interaction with Protein

veFilt	liga	and group	interaction
VOLUL	OH	hydroxyl	hb donor/acceptor
	СО	carbonyl	hb acceptor
	COO	carboxyl	hb acceptor, anion
	COOC	ester	hb acceptor
	COC	ether	hb acceptor
	CON	amide	hb donor/acceptor
	NH	amino	hb donor
	NAR	N aromatic	hb acceptor
	SO3	sulfo	hb acceptor, anion
	SO2N	sulfonamide	hb donor/acceptor
	DON	hb donor	hb donor
	ACC	hb acceptor	hb acceptor
	HAL	halo	halo bond donor
	HPH	hydrophobic	hydrophobic
	STK	aromatic	stacking
	PIC	aromatic	cation $-\pi$



available residue names: ALA, ARG, ASN, ASP, CYS, GLN, GLU, GLY, HIS, ILE, LEU, LYS, MET, PHE, PRO, SER, THR, TRP, TYR, VAL, HOH (ordered water molecule), CA2, MG2, and ZN2 (Ca<sup>2+</sup>, Mg<sup>2+</sup>, and Zn<sup>2+</sup> ions);

> main chain/side chain option;

 $\succ$  minimum number of interactions (default = 1) option



Figure 1. Example of control data used by vsFilt for structural filtration.



# Hydrogen bonds

Maximum distances: upper 90% quantiles of H-bond distances obtained from structural database statistics + 0.5 Å

Bissantz et al. *J. Med. Chem.* (2010)

Liga	and group	Ligand atom	Protein atom	Max distance, Å
			<b>O</b> :	3.5
		hb donor O	eptor O O: 3.5 aromatic N: 3.5 OH: 3.5	
OH	hvdroxvl		aromatic N:	3.5
			OH:	3.5
		hb acceptor O	OH: 3.5 NH: 3.6 OH: 3.5	
CO		11	OH:	3.5
0	carbonyl	nb acceptor O	NH:	3.6
		11	OH:	3.3
		no acceptor O	NH:	3.5
COO	carboxyl		CA2	2.9
		anion O	CA2 2.9 MG2 2.6 ZN2 2.7	
			ZN2	2.7
			OH:	3.5
COOC	ester	hb acceptor O	NH:	3.6
	10C		OH:	3.5
COC	ether	hb acceptor O	NH:	3.6
			<b>O</b> :	3.6
		hb donor N	carboxyl O:	3.5
CON	amide		aromatic N:	3.7
		hh accentor O	OH:	3.5
		по ассерног О	NH:	3.6
			<b>O</b> :	3.6
NH	amino	hb donor N	carboxyl O:	3.5
			aromatic N:	3.7
			OH:	3.5
NAR	N aromatic	hb acceptor N	NH:	3.7
			OH:	3.5
		hb acceptor O	NH:	3.6
SO3	sulfo		CA2	2.9
		anion O	MG2	2.6
			ZN2	2.7
			<b>O</b> :	3.6
		hb donor N	carboxyl O:	3.5
SO2N	sulfonamide		aromatic N:	3.7
		hh accentor O	OH:	3.5
		no acceptor O	NH:	3.6



H-bond angles are constrained to be  $\geq 130^{\circ}$ 



H-bond angles are constrained to be  $\geq 130^{\circ}$ 

Additional vsFilt constraints for H-bonds
Apply angle constraints for h-bonds: O No O Yes
Apply tight constraints for h-bonds: O No 🖲 Yes

Distance ≤ upper 90% quantile (without 0.5 Å increment), angle ≥ 150°



# Hydrogen bonds

DON group can be used as an unspecified H-bond donor and ACC group as an unspecified acceptor:

			O:	3.5–3.6
DON	hb donor	hb donor	carboxyl O:	3.3–3.5
			aromatic N:	3.5–3.7
			Γ	-
	hh accontor	hh accortor	OH:	3.3–3.5
ACC	no acceptor	no acceptor	NH:	3.5–3.7



## **Ionic interactions**

Maximum distance: the mean distance obtained from structural database statistics + 0.5 Å

COO carboxyl	hh accortan O	OH:	3.3
	no acceptor O	NH:	3.5
		CA2	2.9
	anion O	MG2	2.6
			ZN2

SO3 sulfo	hh accentar O	OH:	3.5	
	no acceptor O	NH:	3.6	
	sulfo		CA2	2.9
	anion O	MG2	2.6	
			ZN2	2.7

Zheng et al. J. Inorg. Biochem. (2010)



## Halogen bonds

Maximum distances: upper 90% quantiles obtained from structural database statistics + 0.5 Å

ττατ		Cl	carbonyl O:	3.9
ΠAL	naio	Br, I	carbonyl O:	4.0

Bissantz et al. J. Med. Chem. (2010)



# Hydrophobic interactions

Maximum distances: upper 90% quantiles obtained from structural database statistics + 0.5 Å

			5	
UDU budrochobio	aliphatic C	aliphatic C	4.9	
	anphatie	aromatic C	4.9	
	anomatia C	aliphatic C	4.9	
111 11	при пудгорновіс		aromatic C	4.3
	F	aliphatic C	4.4	
		C1	aliphatic C	4.8

Bissantz et al. J. Med. Chem. (2010)



# Stacking interactions

Maximum distance between centroids of stacked rings:  $\leq$  4.5 Å

STK	aromatic	centroid*	centroid**	4.5
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\*Geometric center of ligand aromatic rings

\*\*Geometric center of Phe/Tyr/His/Trp aromatic rings

Gonzalez et al. J. Phys. Chem. (2000)



# Cation – $\pi$ Interactions

Maximum distances: upper 90% quantiles obtained from structural database statistics + 0.5 Å

PIC aromatic arom	atic C guanidinium C 4.5
-------------------	--------------------------

Bissantz et al. J. Med. Chem. (2010)





vsFilt protein-ligand interaction profile

Provide the interaction profile: 🥐







vsFilt protein-ligand interaction profile Provide the interaction profile: Oreate and edit on-site O Upload as a text file Ligand Protein # of interactions Action OH Add ALA main 🔻 Ŧ 1 Ŧ ALA ۰ funcional group chain Or resnum ARG ASN Add at least one filterin ASP CYS GLN GLU GLY HIS ILE LEU LYS MET PHE PRO SER THR TRP profile (at most 5000 characters) Paste/create/edit your TYR VAL ÷



vsFilt protein-ligand interaction profile

Provide the interaction profile: 🥐

#### Oreate and edit on-site O Upload as a text file



#### Add at least one filtering rule





vsFilt protein-ligand interaction profile

Provide the interaction profile: 🤗

#### Oreate and edit on-site O Upload as a text file

Ligand		Protein		# of interactions	Action
OH 🝷	ALA 🔻		main 👻	1	Add
funcional group	resname	resnum	chain		Or

INTER CON INTER HPH INTER STK	GLY 863 ALA 898 TYR 907	main 2 side side				
Paste/create/	'edit your i	nteraction	profile (at m	ost 5000 cha	racters)	 



# Online analysis

#### Analysis of the vsFilt results

Basic operations with the 3D-viewer; Left-click-and-hold and then move your mouse to rotate the structure, Shift + Left-click-and-hold + Mouse Up/Down to zoom in and out. Ctrl + Right-click-and-hold + Mouse Up/Down/Left/Right to move the structure in the viewer, Right-click for more options. Hold mouse pointer over selected amino acid for one second to view the label. Double click on a selected atom to activate the distance/angle measurement feature. For more refer to the [SMo] manual. Quick hints: Each ligand and its interactions with the protein residues that comply with the filtering rules can be visualized individually by clicking on the respective checkbox, or loaded all at once by using the buttons Toggle all ligands (i.e., all ligands that passed the structural filtration will be shown in the 3D-viewer), Toggle residues (i.e., all residues involved in accommodation of ligands that passed the structural filtration will be shown in the 3D-viewer) and Toggle interactions (i.e., all interactions will be shown between ligands and residues previously enabled in the 3D-viewer). Click on an "cross" icon (🚱) to hide the info for particular ligant (i.e., row) from the table. To restore all rows use the **Restore all rows** button. To highlight a ligand (row) in the table click on any cell (i.e., this feature can help to work with large tables).

#### https://biokinet.belozersky.msu.ru/vsfilt



Viewport: 420x300, 840x600, 1260x900, 1680x1200. Rendering of static image: antialias on (slower), antialias off (faster). Rendering of dynamic image: all features (slower), no antialiasing, no translucency, surfaces dotted, cartoons as trace, geosurfaces as dots, ellipsoids as dots, wireframe only (faster).

Last action with the 3D-viewer:	you have enabled ligand ZINC26894394 ranked #1

Operate PDB Heteatoms:		Show/Hide water		Show/Hide ligands		Show/Hide ions	
Operate Ligands:		Toggle all ligands	l	Toggle residues	Toggle interactions	Restore all rows	
Show/Hide	Rank	Ligand ID	Score	List of interactions			Hide row
	1	ZINC26894394	-9.968	GLY:863 main <mark>O</mark> … N amide	CON		
				GLY:863 main N ··· O amide CON			
				ALA:898 side CB ··· C aromatic HPH			- 8
				ALA:898 side CB ··· C aromat	ic HPH		
			TYR:907 side centroid ··· centro		ntroid aromatic STK		
				GLY:863 main N … O amide	CON		
				GLY:863 main N ··· O amide CON			_
	2	ZINC19522823	-9.364	ALA:898 side CB ··· C aliphat	ic HPH		- 8
				ALA:898 side CB ··· C aliphat	ic HPH		-
				TYR:907 side centroid ··· cer	ntroid aromatic STK		



free (no login required);

can be combined with any type of docking software;

≻implemented using HTML 5;

Processes an SDF library of up to 150 000 docked ligand poses

User data is protected by:

unique access code (TaskID);

> use of HTTPS protocol;

optional IP/password-based authentication

DEMO mode: ~9000 ligands, takes ~2 min





## Illustrative example: PARP ligands

- 236 026 compounds containing a benzamide substructure (classical PARP-1 inhibitor scaffold) from the ZINC12 library;
- Docking with Lead Finder;
- Structural filtration with vsFilt



Figure 1. Example of control data used by vsFilt for structural filtration.



# Illustrative example: PARP ligands

Table 2. Number of Ligands Selected by vsFilt among 236 026 Benzamide Derivatives by Applying Structural Criteria with "Angle Constraints" and "Tight Constraints" Options

interaction	angle constraints	tight constraints	number of ligands
CON…Gly863	-	—	8814
CON…Gly863	+	-	8199
CON…Gly863	+	+	1857
CON…Gly863 HPH…Ala898 STK…Tyr907	+	+	604 (0.26 %)



Figure 1. Example of control data used by vsFilt for structural filtration.

# Illustrative example: PARP ligands





## Illustrative example: PARP ligands



Figure 1. Example of control data used by vsFilt for structural filtration.



**Figure 3.** Interactions of ZINC26894394 ligand with the active site residues in the PARP-1 model: H-bonds with Gly863, hydrophobic contact with Ala863, and  $\pi$ -stacking with Tyr907. The ligand pose was selected by vsFilt and visualized using VMD 1.9.2.<sup>26</sup>

### Пример: 7-Метилгуанин (7-МГ):



- ≻ Метаболит РНК и ДНК
- В небольшой концентрации обнаруживается в моче
- Не используется для синтеза нуклеотидов и не встраивается в ДНК

Weissmann et al. *J. Biol. Chem.* (1957) 224, 407-422 Kaina et al. *Mutat. Res.* (1983) 108, 279-292 Svoboda et al. *Anal. Biochem.* (2004) 334, 239-250

### Пример: 7-Метилгуанин (7-МГ):



- ≻ Метаболит РНК и ДНК
- В небольшой концентрации обнаруживается в моче
- Не используется для синтеза нуклеотидов и не встраивается в ДНК
- Ингибитор ПАРП-1

Weissmann et al. *J. Biol. Chem.* (1957) 224, 407-422 Kaina et al. *Mutat. Res.* (1983) 108, 279-292 Svoboda et al. *Anal. Biochem.* (2004) 334, 239-250 <u>Nilov et al. *Int. J. Mol. Sci.* (2020) 21, 2159</u>

### Биохимические исследования





≻ Конкурентный ингибитор
≻ K<sub>i</sub> ≈ 10 мкМ



**Figure 3.** Dependence of the PARP-1-catalyzed reaction rate on the NAD<sup>+</sup> concentration at different concentrations of 7-MG added to the reaction mixture. Insert: calculated  $K_M^{app}$  values increase with increasing 7-MG concentrations, thus demonstrating the competitive inhibition mechanism.

### Молекулярное моделирование



**Figure 1.** Interactions of 7-MG molecule in the PARP-1 active site revealed by MD simulation: hydrogen bonds with Gly863 and Ser904,  $\pi$ -stacking of purine rings with Tyr907, and hydrophobic contact between the 7-MG methyl group and Ala898.

Нилов и соавт. Acta Naturae (2016) 8, 120-128 Nilov et al. Int. J. Mol. Sci. (2020) 21, 2159 Manasaryan et al. Cancers (2021) 12, 1201



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#### Gushchina et al. J. Chem. Inf. Mod. (2020) 60, 3692-2696







#### vsFilt: A Tool to Improve Virtual Screening by Structural Filtration of Docking Poses

Irina V. Gushchina, $^{\$}$  Aleksandra M. Polenova, Dmitry A. Suplatov, Vytas K. Švedas, and Dmitry K. Nilov $^{\oplus, \vartheta}$ 



ABSTRACT: The ability of ligands to form crucial interactions with a protein target, characteristic for the substrate and/or inhibitors, could be considered a structural filtration of predicted poses improves the performance of virtual screening and helps in recovering specifically bound ligands. Here, we present vsFilt—a highly automated and easy-to-use Web server for postdocking structural filtration. The new tool can detect various types of interactions that are known to be involved in the molecular recognition, including hydrogen and halogen bonds, ionic interactions, hydrophobic contacts, *m*-stacking, and cation-*m* interactions. A case study for poly(ADP-ribose) polymerase I ligands illustrates the utility of the software. The Web server is freely available at https://biolinet.be/ozersky.msur.u/vsfilt.



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